Outlineoffungi.org - Note 994 Ceratobasidiaceae

Web-links: Index Fungorum, Facesoffungi, MycoBank, GenBank

Note: issues around the names *Ceratobasidium* and *Ceratobasidiaceae*

Ceratobasidiaceae Martin (1948) was established based on the genus *Ceratobasidium* D.P. Rogers as typified by the species *C. calosporum* D.P. Rogers (1935). More than 40 species have been described subsequently in *Ceratobasidium*. The family *Ceratobasidiaceae* was treated by Jülich (1982) as also including *Koleroga* Donk, *Oliveonia* Donk, *Scotomyces* Jülich, *Thanatephorus* Donk, *Uthatobasidium* Donk and *Ypsilonidium* Donk. In the 2021 (Outline of Fungi), Wijayawardene et al. (2022) listed the following genera under *Ceratobasidiaceae*: *Ceratobasidium*, *Ceratoporia* Ryvarden & de Meijer, *Ceratorhiza* R.T. Moore, *Rhizoctonia* DC, *Scotomyces* and *Thanatephorus* Donk.

Oberwinkler et al. (2013) examined the holotype of *Ceratobasidium calosporum* and discordant ultrastructural and micromorphological revealed several characters. Ceratobasidium calosporum exhibits partially to completely longitudinally septate basidia, long vermiform basidiospores, and dolipores with continuous parenthesomes. In contrast, all other species described as Ceratobasidium have aseptate basidia, globose to ellipsoid basidiospores, and dolipores with discontinuous parenthesomes (Oberwinkler et al. 2013; Roberts 1999). Oberwinkler et al. (2013) concluded that Ceratobasidium should only be applied to C. calosporum, and the genus was more appropriately placed in the Sebacinaceae K. Wells & Oberw. based on shared characters between C. calosporum and Sebacina calospora (Bourdot & Galzin) Bourdot & Galzin, also known as Ceratosebacina calospora (Bourdot & Galzin) P. Roberts. Based on morphological characteristics, including hyphal diameter and branching pattern along with basidial shape, Roberts (1999) recognized both Ceratobasidium and Thanatephorus, placing a number of genera under Thanatephorus, including Cejpomyces, Aquathanatephorus C.C. Tu & Kimbr., Oncobasidium, Tofispora G. Langer, Uthatobasidium and Ypsilonidium; and placing Koleroga under Ceratobasidium. Roberts (1999) recognized the asexual states of Ceratobasidium and Thanatephorus as Ceratorhiza and Rhizoctonia respectively.

In contrast to the morphological distinction between Ceratobasidium and Thanatephorus, phylogenetic analyses of molecular data have repeatedly demonstrated that there is a single lineage that contains many species described in Ceratobasidium (other than the type) along with the type species of *Rhizoctonia* and *Thanatephorus*, and despite their placement within this lineage, there are not monophyletic groups corresponding to the morphologically-defined genera (Cruz et al. 2022; de Melo et al. 2018; Diederich et al. 2014; Gonzalez et al. 2001; Oberwinkler et al. 2013; Veldre et al. 2013). Thus, it is necessary to recognize a single genus in which Rhizoctonia is the earliest name, and Thanatephorus a synonym, as accepted by Stalpers et al. (2021). Sequence data confirms the synonymy of the following genera under a broadly circumscribed Rhizoctonia: Aquathanatephorus (Vu et al. 2019, under Thanatephorus), Ceratorhiza (Taylor et al. 2003, as "Ceratobasidium goodyerae-repentis", nom. inval.; Xu et al. 2010a), Koleroga (Ceresini et al. 2012, under Ceratobasidum), Oncobasidium (Samuels et al. 2012, under Ceratobasidium) and Uthatobasidium (Roberts 1999, under Thanatephorus; Hibbett & Binder 2002). Morphology remains the rationale for accepting the placement under Rhizoctonia of Cejpomyces, Tofispora and Ypsilonidium. We note that despite the fact that Ceratobasidium is not an appropriate genus for members of the Rhizoctonia clade, new species continue to be described under this outdated generic name (Cruz et al. 2022; de Melo et al. 2018; Diederich et al. 2014).

Because the type species of the type genus of *Ceratobasidiaceae* does not fall in the *Rhizoctonia* lineage, technically the name *Ceratobasidiaceae* cannot be applied to it and the correct family name is *Cejpomycetaceae* Jülich (1982), which was created to accommodate the genus *Cejpomyces* Svrček & Pouzar (1970), a synonym of *Rhizoctonia* (Langer 1994; Oberwinkler et al. 2013; Roberts 1999). However, *Cejpomycetaceae* has been used very rarely — a Google Scholar search gives only two hits compared to more than 5,000 hits for *Ceratobasidiaceae*. A further consequence of the placement of *Ceratobasidium* (in the sense of the type) in *Sebacinaceae*, is that the name for that family should technically be *Ceratobasidiaceae* as this name was introduced in 1948 while *Sebacinaceae* dates from 1982.

It would be highly confusing to replace the widely used family name *Sebacinaceae* with Ceratobasidiaceae, a family name that until recently was used in a quite different sense (i.e. for what is now technically Cejpomycetaceae). In order to retain the use of Ceratobasidiaceae for most species described within it and to avoid replacement of Sebacinaceae, the conservation of Ceratobasidium with a conserved type will avoid disadvantageous nomenclatural changes and a proposal to that end will be submitted shortly. Ceratobasidium sphaerosporum Warcup & P.H.B. Talbot [now Rhizoctonia sphaerospora (Warcup & P.H.B. Talbot) Oberw., R. Bauer, Garnica, R. Kirschner] would be an appropriate type, as there is a sequence available that confirms its position inside the *Rhizoctonia* clade (Gónzalez et al. 2016). Should a proposal to conserve Ceratobasidium with C. sphaerosporum as a type be successful, C. calosporum will need to be placed in another genus. Of the other genera one time included in the Ceratobasidiaceae, Oliveonia is placed in the Oliveoniaceae in molecular phylogeny (Cao et al. 2021), within the Auriculariales J. Schröt. (Cao et al. 2021; Olariaga 2021; Roberts 1998). Reliable sequence data is lacking for Ceratoporia and Scotomyces. Descriptions for these latter two genera state that both genera have clamp connections, while Ceratobasidiaceae is circumscribed as not having clamp connections (Jülich 1982; Roberts 1999). On this basis, Ceratoporia and Scotomyces should be excluded from Ceratobasidiaceae but their systematic position within Cantharellales is uncertain at this time.

References

- Cao T, Hu YP, Yu JR, Wei TZ, Yuan HS. 2021 A phylogenetic overview of the Hydnaceae (*Cantharellales, Basidiomycota*) with new taxa from China. Studies in Mycology 99, 100121. <u>https://doi.org/10.1016/j.simyco.2021.100121</u>
- Ceresini PC, Costa-Souza E, Zala M, Furtado EL, Souza NL. 2012 Evidence that the *Ceratobasidium*-like white-thread blight and black rot fungal pathogens from persimmon and tea crops in the Brazilian Atlantic Forest agroecosystem are two distinct phylospecies. Genetics and Molecular Biology 35(2), 480–497. https://doi.org/10.1590/S1415-47572012005000032
- Cruz EDS, Freitas EFS, Silva MD, Pereira OL, Kasuya MCM. 2022 A new mycorrhizal species of *Ceratobasidium (Ceratobasidiaceae)* associated with roots of the epiphytic orchid *Gomesa recurva* from Brazilian Atlantic Forest. Phytotaxa 550(3), 224–32. https://doi.org/10.11646/phytotaxa.550.3.2
- de Melo MP, Matos KS, Moreira SI, Silva FF et al. 2018 Two new *Ceratobasidium* species causing white thread blight on tropical plants in Brazil. Tropical Plant Pathology 43(6), 559–571. <u>https://doi.org/10.1007/s40858-018-0237-x</u>
- Diederich P, Lawrey J, Capdet M, Pereira S et al. 2014 New lichen-associated bulbilforming species of Cantharellales (*Basidiomycetes*). The Lichenologist 46, 333–347. <u>https://doi.org/10.1017/S0024282913000583</u>

- Gonzalez D, Carling, DE, Kuninaga S, Vilgalys R,Cubeta MA. 2001 Ribosomal DNA Systematics of *Ceratobasidium* and *Thanatephorus* with *Rhizoctonia* anamorphs. Mycologia 93(6), 1138–1150. https://doi.org/10.2307/3761674
- Gónzalez D, Rodriguez-Carres M, Boekhout T, Stalpers J et al. 2016 Phylogenetic relationships of *Rhizoctonia* fungi within the Cantharellales. Fungal Biology 120(4), 603–619. <u>https://doi.org/10.1016/j.funbio.2016.01.012</u>
- Hibbett DS, Binder M. 2002 Evolution of complex fruiting–body morphologies in homobasidiomycetes. Proceedings of the Royal Society of London. Series B: Biological Sciences 269(1504), 1963–1969. <u>https://doi.org/10.1098/rspb.2002.2123</u>
- Jülich W. 1982 Higher Taxa of Basidiomycetes (Vol. 85). J. Cramer.
- Langer G. 1994 Die gattung Botryobasidium Donk (*Corticiaceae, Basidiomycetes*). Schweizerbart Science Publishers. <u>http://www.schweizerbart.de//publications/detail/isbn/9783443590604/Bibliotheca_M</u> ycologica_Band_158
- Martin GW. 1948 New or noteworthy tropical fungi IV. Lloydia, 11, 111–122.
- Oberwinkler F, Riess K, Bauer R, Kirschner R, Garnica S. 2013 Taxonomic re-evaluation of the *Ceratobasidium-Rhizoctonia* complex and *Rhizoctonia butinii*, a new species attacking spruce. Mycological Progress 12(4), 763–776. https://doi.org/10.1007/s11557-013-0936-0
- Olariaga I. 2021 Cantharellales Gäum. In Ó. Zaragoza & A. Casadevall (Eds.), Encyclopedia of Mycology (pp. 320–328). Elsevier. <u>https://doi.org/10.1016/B978-0-12-819990-9.00056-1</u>
- Roberts P. 1998 *Oliveonia* and the origin of the *holobasidiomycetes*. Folia Cryptogamica Estonia 33, 127–132.
- Roberts P. 1999 *Rhizoctonia*-forming Fungi: A Taxonomic Guide. Herbarium, Royal Botanic Gardens Kew.
- Rogers DP. 1935 Notes on the lower basidiomycetes. Studies in Natural History, Iowa University, 17(1), 1–43.
- Samuels GJ, Ismaiel A, Rosmana A, Junaid M et al. 2012 Vascular Streak Dieback of cacao in Southeast Asia and Melanesia: In planta detection of the pathogen and a new taxonomy. Fungal Biology 116(1), 11–23. https://doi.org/10.1016/j.funbio.2011.07.009
- Stalpers JA, Redhead SA, May TW, Rossman AY et al. 2021 Competing sexual-asexual generic names in *Agaricomycotina (Basidiomycota)* with recommendations for use. IMA Fungus 12(1), 22. <u>https://doi.org/10.1186/s43008-021-00061-3</u>
- Svrček M, Pouzar Z. 1970 *Cejpomyces* gen. nov., a new genus of resupinate *Hymenomycetes* (*Corticiaceae*). Česka Mykologie 24(1), 5–11.
- Veldre V, Abarenkov K, Bahram M, Martos F et al. 2013 Evolution of nutritional modes of *Ceratobasidiaceae* (*Cantharellales*, *Basidiomycota*) as revealed from publicly available ITS sequences. Fungal Ecology 6(4), 256–268. https://doi.org/10.1016/j.funeco.2013.03.004
- Vu D, Groenewald M, de Vries M, Gehrmann T et al. 2019 Large-scale generation and analysis of filamentous fungal DNA barcodes boosts coverage for kingdom fungi and reveals thresholds for fungal species and higher taxon delimitation. Studies in Mycology 92, 135–154. <u>https://doi.org/10.1016/j.simyco.2018.05.001</u>
- Wijayawardene N, Hyde K, Dai DQ, Sanchez-Garcia M et al. 2022 Outline of Fungi and fungus-like taxa – 2021. Mycosphere 13, 53–453. https://doi.org/10.5943/mycosphere/13/1/2

Xu Z, Harrington TC, Gleason ML, Batzer JC. 2010 – Phylogenetic placement of plant pathogenic *Sclerotium* species among teleomorph genera. Mycologia 102(2), 337–346. https://doi.org/10.3852/08-189.

Entry by

Ryan P. O'Donnell Division of Ecology & Evolution, Research School of Biology, Australian National University, Canberra ACT, 2612, Australia **Tom W. May** Royal Botanic Gardens Victoria, Melbourne VIC 3004, Australia

(Edited by: Kevin D. Hyde & Maryam Tavakol Noorabadi)

Published online 5 April 2024